**Cytometer Setup and Tracking (CST)**

1. CST beads preparation:   
   Choose the right cst stock (Blue -Fortessa and Aria new; Orange- Aria old-)  
   Shake the stock. Put one drop in 350µl PBS. Vortex.
2. Change density filter to 1.0. Close the lid.
3. In the Diva software: cytometer 🡪CST
4. A new workspace will appear. If you don't see it minimized the Diva window.
5. Choose lot id.
6. Run performance. Answer the questions.
7. It will run for ~10min.
8. Review the report  
   **OK**- perfect ☺ close and go back to your exp.

**!** – passed with remarks. Open the report and check which channel failed.

**Fail** – take a photo of the message. Report to the person in charge. Make sure the right density filter is

**Clean flow cell**

When we use clean flow cell with contrad: if in the cst report there are indications for distorted scatter or high rCVs.

1. Prepare 1:5 contrad solution in water.
2. **Stop the stream. Remove the nozzle and defection plates - do not perform with the nozzle inside!**
3. Cytometer-🡪 cleaning mode 🡪 clean flow cell X3 after third time – wait at least 5 min (the longest the better).
4. Activate stream – there is no nozzle so there will be no drops.
5. Move to any experiment. Load an empty tube, flow rate 11, 3-5 min. stop🡪 unload.
6. Close stream, dry flow cell and plates with kimwipes and sticks, don't touch with your hands. Dry well.
7. Insert the correct nozzle and put back the plates.
8. Activate the stream – make sure the drops look stable. Increase/decrease amp if needed.

Clean flow cell with **clean solution** is also optional. Recommended after running yeasts or dyes like PI that tend to stick. Use the cleaning nozzle. 5 min incubation are enough. To wash out the clean solution 🡪 3x clean flow cell on water.

If you want to check if there is a clog in the sample line you can use **Clean flow cell** on water just to see if the water in the tube is getting down.